BIOSYNTHESIS OF PH RESPONSIVE SHAPE MEMORY HYDROGEL AND ITS BIOMEDICAL POTENTIAL

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Abstract

Multifunctional hydrogels combining the capabilities of cellular pH responsiveness and shape memory, are highly promising for the realization of smart membrane filters, controlled drug released devices, and functional tissue-engineering scaffolds. Free radical biocatalytic polymerization catalyzed by immobilized Candida antarctica lipase B was used to fabricate the pH-responsive and shape memory hydrogel using medium-chain-length poly-3-hydroxyalkanoates-co-polyethylene glycol methacrylate (PHA-PEGMA) as macromer. The accelerative wound healing potential of the biosynthesized smart PHA based hydrogel was evaluated herein. The thermal stability of the macromer highly depends on the PEGMA fraction from 10 to 50% (mass). Similarly, the change in PEGMA fraction was also found to highly influence the hydrogel's hydration rate (h) from 2.83 x 10^5 to 7.63 x 10^5 mL/s. The hydrogel's equilibrium weight swelling ratio (qₑ), protein release and its diffusion coefficient (Dₘ) were all found to be pH dependent. For example, increasing the phosphate buffer pH from 2.4 to 13 resulted in increased qₑ from 2 to 16 corresponding to the enlarging of network pore size (ξ) from 150 to 586 nm. The biomedical potential of the synthesized material based on its accelerative wound healing in rats was observed. Biochemical, histomorphometric and immunohistomorphometric analyses revealed a significant difference in area closure and re-epithelialization on days 7 and 14 in PPH or Intrasite® gel group compared to gum acacia or PEGMA-alone groups. Furthermore, wounds dressed with PPH or Intrasite® gel showed evident collagen deposition, enhanced fibrosis and extensively organized angiogenesis on day 14 compared to the negative control group. The findings suggested that topical application of PPH accelerated the rats’ wound healing process by improving angiogenesis attributed to the increased microvessel density (MVD) and expressions of VEGF-A in tissue samples. Thus, PPH has been demonstrated to be effective in the treatment of cutaneous wounds in rats, and could be a potential novel agent in the management and acceleration of wound healing in humans and animals.

Key Words: Biocatalysis, Biomaterials, Biototechnology, Hydrogel, Wound-Healing.

Introduction

The effective control and treatment of some cutaneous wound under certain physiological or pathological conditions is problematic. This is due to the factors that interfere with the sequence of the wound healing process, leading to a delayed or impaired wound healing response, culminating in a revolting scar. As a result of these complications, impaired wound healing is an expensive and time-consuming undertaking. Inadvertently, it brings sociological, economical and public health burdens on the patients. Consequently, exploring affordable, effective and readily accessible alternative therapies for wound healing is currently an active area of research. Among such approaches are the use of phytochemical extracts (Al-Bayaty and Abdulla 2012, de Oliveira et al. 2013), carbohydrate based polymer composite (Nguyen et al. 2013).

Along this line, the biodegradability and biocompatibility of natural polymers such as poly-3-hydroxy alkanoates (PHAs) with the advantage of being produced by bacterial fermentation from renewable resources, or by in vitro green enzymatic catalysis strongly favor their applications in smart actuators, smart membrane filters, controlled drug released devices and tissue-engineering scaffolds etc.

Hydrogels made of amphiphilic polymer composites containing the biodegradable hydrophilic and lipophilic PHA are interesting due to their environment-sensitive micellar properties, such as temperature and pH responsiveness, excellent biocompatibility, lack of pro-inflammatory effects of polymeric systems and controlled capacity in protein release (Hennink et al. 1997). Since their first description by Guiseppi-Elie and Sheppard (1995), hydrogels are being exploited in several potential applications such as site-specific drug delivery devices (Schoener et al. 2012). In fact hydrogels made from biocompatible polyhydroxyalkanoates (PHA), polyanide and/or polyamino acids composites were reported to exhibit biomimetic properties especially in bioactuators, biosensors, smart ion exchange membranes, cell-adhesion or targeting, and their efficacy is well distinguished in cancer biology and innate immunity (Suresh 2007).

The use of PHAs based hydrogels in accelerative cutaneous wound healing presents several advantages over the commonly used alternatives. For example, the diverse structural composition of PHAs confers myriad of superior physico-mechanical properties. These include crystallinity, flexibility, thermal stability, piezoelectricity, porosity etc. Owing to these properties, the neat polymer or its composite could be suitably casted as film and used as superb adhesive bandage. The bioactivity of polyhydroxy- group has been reported Teramoto et al. (2008). Since the PHA backbone is adorned with numerous such groups, an inherent bioactivity could be perceived.

Despite their reported biomedical importance spanning from antimicrobial (Saad et al. 2012), target-guided devices (Yao et al. 2008), controlled drug release devices (Xiong et al. 2010), smart hydrogel (Gumel and Annuar 2014), to surgical implant (Türesin et al. 2001), research on the use of biocompatible and biodegradable PHAs in accelerative wound healing is very scarce (Bai et al. 2014).
In this study, mild-conditioned dual enzymatic catalysis was used to prepare a macromer and its respective smart hydrogel. The characterization of the said biomaterial were also studied and it accelerative wound healing activity was observed.

2.2 Methods

2.2.1 Enzymatic synthesis of poly(3-hydroxyalkanoates)-co-(6-hydroxyhexanoate) macromer (PHA-PCL)

Bacterial mcl-PHAs was obtained according to previously reported methods (Gumel et al. 2012). The PHA sample (200 mg) was dissolved in a capped vial containing 10.0 ml toluene. To this solution, 3.0 ml of ε-caprolactone and 150 mg of immobilized enzyme (Novozym) were added. Then, the reaction mixture was incubated in an automatic orbital shaker (Labtech, Korea) at 50 (±1) °C, 250 rpm for 48 hours.

At the end of the reaction, the synthesized macromer was then extracted from the crude reaction mixture as reported somewhereelse (Gumel and Annuar 2014).

2.2.2 Synthesis of the poly(3-hydroxyalkanoates)-co-(6-hydroxyhexanoate) based hydrogel

The hydrogel film was synthesized by free radical polymerization as illustrated in Scheme 1. A solution of PHA-PCL macromer was prepared by dissolving the macromer (100 mg) in 1.5 ml chloroform and mixed with 1.0 ml solution of 20 % (w/w) PEGMA in 50 mM physiological phosphate buffer (pH 7). PEGMA acted as a cross-linker. The mixture was sparged with nitrogen gas for 5 min. 120 µl of freshly prepared solutions of 10 % w/v APS and 100 µl of 20 % v/v TEMED that served as free radical initiators were sequentially added to the mixture. Following this, the reaction mixture was quickly poured into a 10 ml beaker (2.5 cm internal diameter) at 2 mm thickness and allowed to polymerize for 20 min at 25 °C. Thereafter, the synthesized hydrogel was freed out of impurities by sterilized water immersion followed by oven drying prior to experimental testing (Figure 1). Characterization of the synthesized biomaterial was observed as reported previously (Gumel et al. 2015).

2.2.6 Experimental Animal Model

The accelerative wound healing activity of PHA-PCL based hydrogel film was evaluated using rats model as reported in literature (Hajiaghaalipour et al. 2013). The experimental wound excision, wound dressing and wound area contracture were all observed according to literature (Kirker et al. 2002).

2.2.7 Wound Excision

The initial weight of each rat was recorded before the infliction of wound excision. After the weighing, the experimental wound excision was inflicted on the rat as described in literature (Al-Bayaty and Abdulla 2012).

2.2.10 Immunohistochemical analysis

The immunohistochemical staining for the determination of microvessel density (MVD) and expression of vascular endothelial growth factor A (VEGF-A) was performed according to the method of Zhang et al. (2010)

2.2.11 Wound tissue homogenate protein content

The total protein content of the wound tissue from each animal was determined according to the method described by Al-Bayaty and Abdulla (2012).
2.2.12 Estimation of connective tissue parameters

Tissue samples collected for the estimation of connective tissue parameters (hydroxyproline, hexosamine and hexuronic acid contents) were prepared according to the methods explained in literature (Murthy et al. 2013).

2.2.12.1 Hydroxyproline (HPR), Hexosamine (HXA) and Hexuronic acid (HUA) estimation

Wound tissue hydroxyproline content was estimated following the method described by Murthy et al. (2013) and (Gumel et al. 2015).

2.3 Statistical analysis

Statistical analysis was performed using Sigmaplot ver. 11.0 (SYSTAT™, USA). All experimental data were presented as average ± standard deviation and statistical significance were determined using ANOVA at 95% confidence interval.

3. Results

3.1 In vitro cytotoxicity assay

Human HeLa cell derivatives (WRL68) was used as a cellular model during the evaluation of the hydrogel’s cytotoxicity following MTT reduction assay. With reference to control sample, the results demonstrated that increasing the hydrogel sample size up to 2.5 g did not caused a significant decrease in cell viability even after 72 hours incubation (96 % cell viability at 2.5 g hydrogel extract).

3.3 Percentage wound contraction

The percentage of wound contraction was measured at day 3, 7 and 14. At the early phase of the wound healing (day 3), there was no significant difference between the negative control group and the treated groups. On day 7, the wound area in the whole samples was found to contract in comparison to the initial excision time. However, no significant difference was observed between the negative control group (10 %) and the secondary control (PH) group (13 %). In contrast to the negative control group, there was a significant difference (p < 0.05) in wound contracture of PPH (49 %) and Intrasite® (56 %) groups. As the wound aged to day 14, the PH dressed group was observed to show marked increase in wound contraction (45 %) when compared to the gum acacia treatment group (30 %). At this time, the increase was found to be significantly different (p < 0.05) in both PPH (76 %) and Intrasite® dressed groups (80 %). It could be observed that PPH dressing showed comparable wound contraction performance to Intrasite® gel.

3.5 Immunohistochemical analysis

On day 14 post-excision, the MVD and the expression of VEGF-A were found to be at peak for all the sample tested (Figure ). However, with reference to negative control (Figure 4 (a) and (e)) and PH dressed (Figure 4(b) and (f)) groups, the PPH dressed group revealed a significant (p < 0.05) increased in both MVD and VEGF-A expression (Figure 4 (c) and (g)). In fact, the immunohistomorphometric appearance of the PPH dressed micrographs indicated a normal wound healing process that was accompanied by high angiogenesis and VEGF-A expression to a level almost similar to that observed in the micrographs of the standard Intrasite® gel dressed group (Figure 4 (d) and (h)).

![Figure 2](https://cirworld.com)

Figure 2. Immunohistochemical micrographs of excisional wound tissue on day 14 of dressing stained with anti-CD31 (a-d) and anti-VEGF-A (e-h) antibodies at 400 × magnification: (a) negative control (b) PH dressed group; (c) PPH dressed group; (d) Intrasite dressed group. VEGF-A stain (e) negative control group; (f) PH dressed group; (g) PPH dressed group and (h) Intrasite® gel dressed group. The red arrow depicts micro vessel density and yellow arrow denotes the VEGF-A expression.
3.6 Tissue homogenates protein content

The total protein content was found to significantly increase (p <0.05) in both the PPH (276 ±10.4 mg/ g tissue homogenates) and Intrasite® gel (308 ±12.3 mg/ g tissue homogenates) dressed groups as compared to the negative control group (Figure 6). In contrast, although there was a marked increase in protein content of PH dressed group (208 ±14.2 mg/ g tissue homogenates), it was not significant compared to the negative control group (189 ±19.3 mg/ g tissue homogenates). In general, higher protein level comparable to that of Intrasite® gel dressed group was observed in wounds dressed with PPH (Figure 6).

3.7 Connective tissue parameters

With reference to the data for negative control group, the HPR, HXA and HUA values were significantly (p < 0.05) increased in PH, PPH and Intrasite® dressed groups, respectively (Table 1). In comparison to the gum acacia control group, the HPR, HXA and HUA were significantly increased in PPH dressed group by 42 %, 59 % and 163 %, respectively. Murthy et al. (2013) reported a similar increase in HPR, HXA and HUA when investigating the wound healing activities of Bacopa monniera in rats.

Table 1 Connective tissue parameters on the effects of PHA-PCL hydrogel dressing on excisional wound healing

<table>
<thead>
<tr>
<th>Dressing</th>
<th>Hydroxyproline (µg/mg tissue hydrolysate)</th>
<th>Hexosamine (µg/mg tissue hydrolysate)</th>
<th>Hexuronic acid (µg/mg tissue hydrolysate)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative control</td>
<td>148.9 ± 7.4</td>
<td>78.2 ± 2.9</td>
<td>19.5 ± 1.2</td>
</tr>
<tr>
<td>PH dressed</td>
<td>172.3 ± 8.6</td>
<td>98.3 ± 5.8</td>
<td>24.5 ± 2.4</td>
</tr>
<tr>
<td>PPH dressed</td>
<td>212.1 ± 12.2</td>
<td>124.4 ± 6.2</td>
<td>51.8 ± 2.6</td>
</tr>
<tr>
<td>Intrasite® gel</td>
<td>228.6 ± 11.3</td>
<td>127.5 ± 4.6</td>
<td>53.4 ± 2.7</td>
</tr>
</tbody>
</table>

Data values are expressed as average ± standard deviation of six (6) rats per each group. An asterisk (*) represents a significant (p < 0.05) difference when compared to negative control group.

4. Discussions

In this study, we evaluated the effects of topical dressing with chemo-enzymatically synthesized hydrogel based on biocompatible PHA-PCL composite on skin excision wounds in rats. In this research, in vitro MTT cytotoxicity evaluation revealed no significant difference (p > 0.05) in terms of WRL68 cellular proliferation and viability between the control group and the leachable residues of the composite polymer hydrogel samples even at longer exposure time (72 hours) and higher material weightage (2.5 g). This observation was found to be in agreement to other similar observations of high cellular viability (> 80%) following hydrogel cytotoxicity tests (Draye et al. 1998, Trudel and Massia 2002, Wu et al. 2008).

Histological examination of the excisional wound area revealed an observable increase in wound repair upon topical dressing with the synthesized hydrogels. Wound contraction was found to occur in the early days of post wound excision upon topical dressing. A dramatic increase in wound contraction was observed to occur within 7 to 10 days post excision. In fact, a significant increase (p < 0.05) in the percentage of wound contraction was observed in PPH and Intrasite® gel dressed groups on day 7 of post excision and the area was further reduced on day 14 post excision. Related decrease in the percentage of wound contracture upon dressing with aminated and chlorine dioxide gels was reported in streptozotocin-induced diabetic rats (Al-Bayaty and Abdulla 2012). Similarly, significant increase in wound contracture was observed on day 7 in thermosensitive polyethylene glycol triblock hydrogel (Lee et al. 2003). In another study, topical dressing with glycosaminoglycan hydrogel revealed no significant difference between the control group and the treated groups (Kirker et al. 2002). However, by day 7 post excision, the authors reported a significant increase in re-epithelialization (Kirker et al. 2002). Evidently, in this study, the increase in wound contracture corroborated with a marked increase in re-epithelialization in groups dressed with PPH and Intrasite® gels. In addition to re-epithelialization, the excision wounds revealed faster healing process due to the observed extensive increase in granulation tissue and reduced number of mononuclear inflammatory cells in these groups. Furthermore, a reduction in eosinophil infiltration, increased neovascularization, restoration of adnexa, and pronounced fibrosis was also noted.

The micrographs obtained from the Mason’s trichrome stain further supported these observations due to the presence of elevated levels of angiogenesis and collagen deposition. Angiogenesis increases the delivery of much need oxygen and nutrients for cellular proliferation and collagen deposition within the wound area. Impaired wound healing process is attributed low blood flow, which consequently resulted in lower oxygen supply and poor collagen deposition (Al-Bayaty and Abdulla 2012). However, influx of the macrophages and the increased neovascularization accompanied by strong collagen deposition in the granulation tissue could explain the mechanism underlying the topical dressing of PPH and Intrasite®
Cytokines are low molecular weight proteins (~4 to 60 kDa) and function mostly in cellular signaling (Diegelmann and Evans 2004). It has been hypothesized that cytokines achieved the function of cell signaling by altering the intracellular kinase activity and phosphorylation of major proteins consequently altering the cellular physiological status and cellular function (Debono 2000). Armed with cellular signaling ability, cytokines were reported to be the molecular time controlled switch between different phases of wound healing process (Debono 2000). During these processes, several cytokines activities are known to take place e.g. transforming growth factor beta (TGF-β) is known to be involved in wound healing and scar formation (O’Kane and Ferguson 1997). Similarly, platelet-derived growth factor (PDGF) that is attributed to pro-inflammatory cytokine protein of dimeric structure attracts leucocytes and fibroblasts to the healing wound (Slavin 1996). Furthermore, vascular endothelial growth factor (VEGF) which is also a dimeric protein and a heparin-binding endothelial growth factor is delineated to be specifically associated with vascular endothelial cells (Slavin 1996). Its binding sites on vascular endothelium were previously illuminated (Ferrara et al. 1991). In fact, in situ molecular hybridization have revealed the VEGF mRNA to be highly expressed and abundantly distributed in the areas of active vascular proliferation (Slavin 1996). Recent studies have reported a marked reduction in pig's wound angiogenesis, fluid accumulation, and granulation tissue formation upon application of neutralizing VEGF-A antibodies (Howdieshell et al. 2001, Werner and Grose 2003). Moreover, neutralization of VEGF was demonstrated to strongly inhibit angiogenic activity in human wound (Nissen et al. 1998). Finally, a pronounced reduction in angiogenesis and granulation tissue development was observed in murine cutaneous wound upon delivery of retroviral dominant-negative VEGFR-2 (Tsou et al. 2002).

In this study, we evaluated the microvessel density (MVD) and the expression of VEGF using immunohistochemical staining. As expected, with reference to the control and PH dressed groups, there was an observed marked increase in MVD and VEGF-expression in both PPH and Intrasite® gel dressed samples. This clearly acceded to the prior literature reports and our own observation of neovascularization in H&E staining micrographs of the same samples. Similar report on enhanced wound healing process due to increased MVD and elevated expression of VEGF-A was reported (Kirker et al. 2002). During wound healing process, lipids and fatty acids are postulated to play an important role in the development of new micro-vessels and elevated VEGF-A expression (Sellmayer et al. 1999, Kirker et al. 2002, Claria 2006). Previously, the bioactivity of polyhydroxy- compounds including PHA and PCL was established (Teramoto et al. 2008, Gumel et al. 2013). Since PPH is based on PHA-PCL copolymers composed of 3-hydroxy and 6-hydroxy fatty acids monomers, respectively, this could probably explain the underlying mechanism for the observed increase in MVD and elevated VEGF-A expression in PPH dressed group.

Conclusion

Enzymatically synthesized PHA-PCL macromers were cross-linked with PEGMA to give a bioactive hydrogel that can be aseptically sterilized and topically applied on cutaneous wound in experimental rat models. Significant difference in area contraction and pronounced re-epithelialization was found on day 7 and 14 in the wounds dressed with the PPH and Intrasite® gel compared to those dressed with gum acacia or PEGMA alone. Additionally, those wounds dressed with PPH or Intrasite® gel showed marked collagen deposition, more fibrosis and extensively organized angiogenesis by day 14 compared to negative control or PH dressed groups. There were no significant differences between the negative control group and the PH dressed group on any day post wound-excision.

The findings suggested that topical application of the PPH helped to accelerate experimental rats wound healing by improving angiogenesis, which was attributed to the increased in MVD and expression of VEGF-A in tissue samples. Thus, PPH has been demonstrated to be effective in the treatment of cutaneous wounds in rats. It could be potentially employed as a novel agent for the management and acceleration of wound healing in human and animals.

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Conflict of Interest
The author declares that there is no existence of competing interests, financially or otherwise.

1. References


