

Colloidal Quantum Dots Suitability for Long Term Cell Imaging

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Abstract

Fluorescent semiconductor nanoparticles in tree-dimensional quantum confinement, quantum dots (QDs), synthesized in aqueous medium, and functionalized with polyethylene glycol, were used as nonspecific nanosized probes for the long-term imaging of glial cells. *In vitro* living healthy as well as cancer glial cells were labelled by direct insertion of a small volume of QDs contained in aqueous suspension into the culture wells. A long-term monitoring (over 7 days) of the cells was performed and no evidence of cell fixation and/or damage was observed. Two control groups, healthy and non-healthy glial cells, were used to compare cell viability. During the observation period, labelled cells kept the same behavior compared to non-labelled control samples. To our knowledge, this is the first report of the viability of quantum dots for long-term imaging of living cells. This opens a large range of possibilities related to a better understand of cellular mechanisms, which till now was not achieved by any other fluorescent probe. The study also permitted the observation of two distinct interaction kinetics between cells and QDs. Healthy glial cells were mainly labelled at their surface, while non-healthy glial cells have shown a high rate in the uptake of QDs.

Key-words: quantum dots, nanoprobles, long term cell imaging, bioimaging, fluorescence microscopy.

1- Introduction

When light interacts with semiconducting nanoparticles, valence band electrons may acquire enough energy to bridge the energy gap and reach the conduction band. The excited electrons may decay to their original state by means of radiative and/or non-radiative mechanisms [1,2]. The radiative ones gave rise to many fluorescence-based assays, while the non-radiative have been used increasingly in photoacoustic (PA) [3,4]. These two classes of mechanisms are in particular present and their physicochemical properties, which may be selected by tuning their size. In these cases, the electrons of each particle are confined in each one of the 3 dimensions. Nanoparticles under such conditions constitute examples of low dimensional systems. In such a case, a class of zero dimensional (0D) materials under quantum confinement regime in all the 3 dimensions. These systems are named quantum dots (QDs) [5].

In vivo and *in vitro* toxicological studies of QDs have been carried out during the last decade [6-10]. For *in vitro* diagnostic assays, such as fluorescence microscopy (single and multi-photon), QDs have shown to be very efficient, even surpassing conventional dyes, once QDs allow stoichiometrically prepared surfaces for specific targeting in cells and tissue samples, without the need of fixation of such samples. This feature opens entirely new possibilities for bio-imaging based diagnostics techniques. To qualify as probes for *in vitro* diagnostics, QDs must be biocompatible. This condition may be achieved, both for bottom-up [11-13] and top-down [14-16] prepared QDs, by modifying their surface *via* the controlled addition of layers of organic polyelectrolytes, peptides and/or other biomolecules, such as mono or polyclonal antibodies [17]. Migration of Glioblastoma Multiform (GBM) cells has been pointed out as a crucial factor underlying the aggressive nature of GBM [18] and its infiltration in central nervous system. Fluorescent probes that allow long term *in vitro* observation of GBM cells, must present low or null toxicity for the cells and may provide significant improvements for brain cancer theranostic approaches.



2- Methodology

Otherwise mentioned, chemicals were purchased from Sigma-Aldrich and used without further purification. The C6 rat glioma cells and the healthy glial cells (rat astrocytes) were purchased from Sigma-Aldrich for research purposes only, and are part of the European Collection of Authenticated Cell Culture (ECACC), as declared by Sigma-Aldrich. This way, ethical issues are covered by the certification of ECACC. Core-shell Cadmium Sulphide/Cadmium Hydroxide CdS/Cd(OH)₂ QDs (4nDOT500) designed for a peak emission of 500 nm were produced by Phornano Holding GmbH, Korneuburg, Austria. The passivation step of CdS nanoparticles consisted of a controlled precipitation of Cd(OH)₂ layers at high pH medium. QDs were characterized by electronic absorption and emission spectroscopies, conventional and high-resolution transmission electronic microscopy (HR-TEM and TEM) and X-ray diffractometry (XRD). 4nDOT500 are functionalized by polyethylene-glycol (PEG) (molecular weight: 400). QDs were chemically associated to PEG in aqueous media using 0.1M PEG aqueous solution (0.0025 g/mL) at room temperature (RT) at pH 7.2. QDs were centrifuged and two main sizes population were identified: 6 and 9 nm. Rat C6 glioma cells were kept in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin at 37°C in a 5% CO₂ atmosphere. The C6 cells were seeded at 5×10⁴ cells/well in two sets of 96-well microplates containing DMEM. The cell sets were then named as *L* and *C* (*L* = *Labelled* and *C* = *Control*). Both sets *L* and *C* were analyzed within the same acquisition parameters. Prior to the visualization at the confocal fluorescence microscope set-up, the cells of *L* set were washed with PBS and then, directly visualized by laser scanning confocal microscopy (Leica Microsystems GmbH, Wetzlar, Germany) with an apochromatic water-immersion objective (100X, NA 1.2) at wavelengths of 488 and 533 nm. Fluorescence images presented in next section are the ones resulting from overlaying images obtained by exciting samples at the two wavelengths mentioned above (488 and 533 nm). The same procedure was used for the analysis of control cells (set *C*). Confocal microscopy and viability assays were performed at different intervals during 7 days in order to monitor long-term cell viability and to detect possible morphological damage of the cells. All images were taken with the same acquisition parameters and processed using modular imaging software for fluorescence imaging (Leica QWin Lite, Leica Microsystems GmbH, Wetzlar, Germany) for performing all the assays. Cell viability was then determined using an EZ-Cytox Cell Viability Assay kit according to the manufacturer's protocol.

The same core/shell Cadmium Sulfide/Cadmium Hydroxide QDs in aqueous medium used in this work, have been previously analyzed by means of *in vivo* toxicological studies using Male Wistar rats, which were head-nose exposed to a liquid aerosol of the Cd-based QDs, 6 hours per day on 5 consecutive days [19,20].

3. Results

Emission spectrum obtained for as prepared QDs/PEG is presented in Figure 1 (bottom). No fitting algorithm or mathematical post processing has been applied. The maximum emission intensity occurs at 504 nm. In Figure 1 (top), the experimental and the theoretical (vertical lines) X-ray Diffractogram are shown. By analyzing XRD it can be observed that the CdS/Cd(OH)₂ QDs present cubic crystalline structure, which corresponds to the more stable crystalline form for CdS [21].

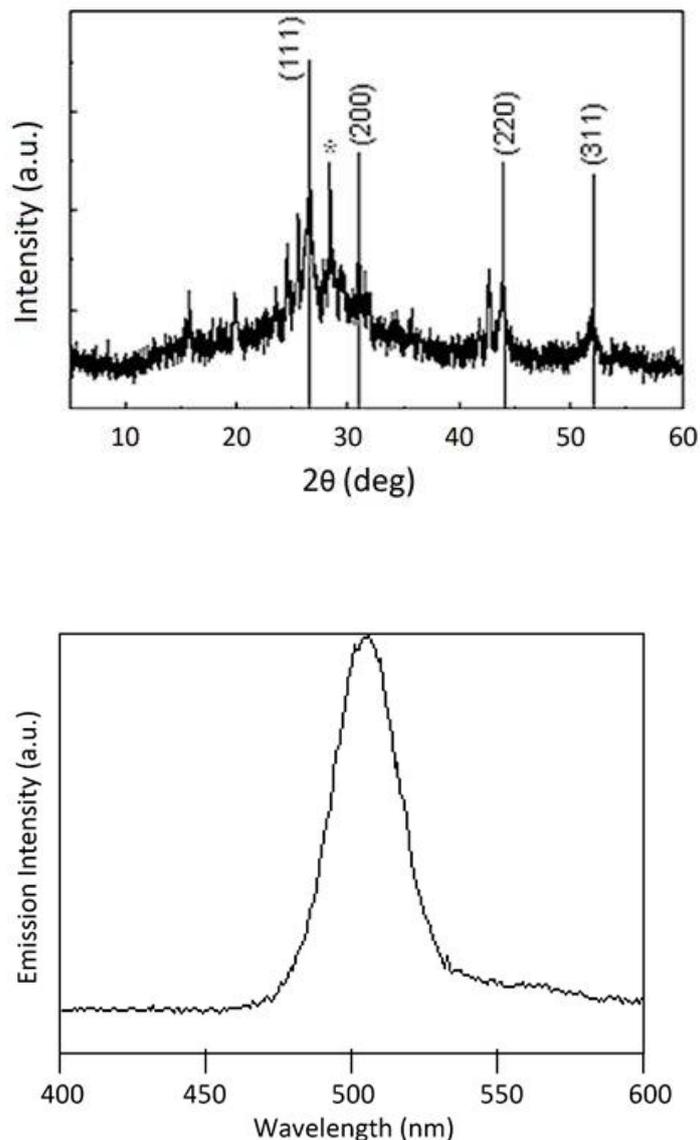


Figure 1-Top: Experimental diffractogram and theoretical (vertical numbered lines) XRD for the QDs. Vertical lines correspond to cubic faces peaks. **Figure 1 -Bottom:** Emission spectrum of as prepared pegylated QDs (4nDOT500)

Cell viability results are presented in Figure 2. No significant differences between labelled (*L* set) and non-labelled (*C* set) glioblastomas were observed.

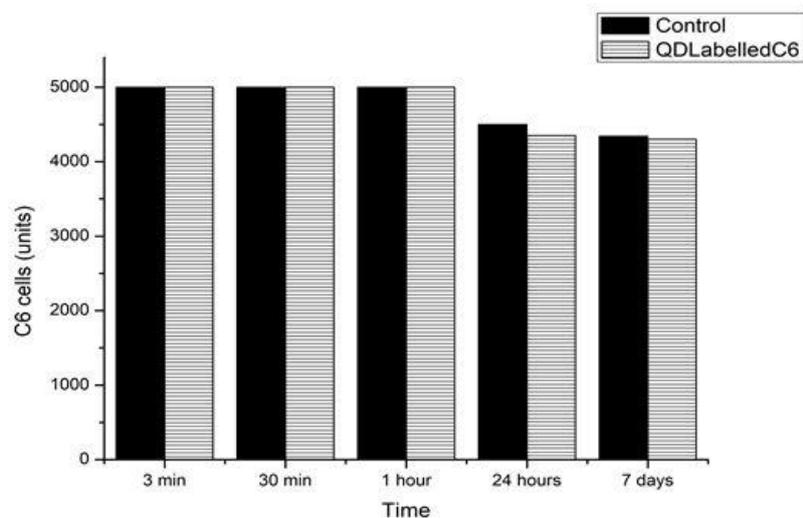


Figure 2: Cell viability for non-labelled (Control) C6 glioblastoma cells and for QD-labelled C6 glioblastoma cells. The numbers are the average obtained for the 96 wells of each set.

Fluorescence image obtained by single photon Laser Scanning Confocal Microscopy for labelled C6 glioblastoma cells, with incubation times of 3 minutes as well as 7 days are shown in Figure 3 and Figure 4, respectively.

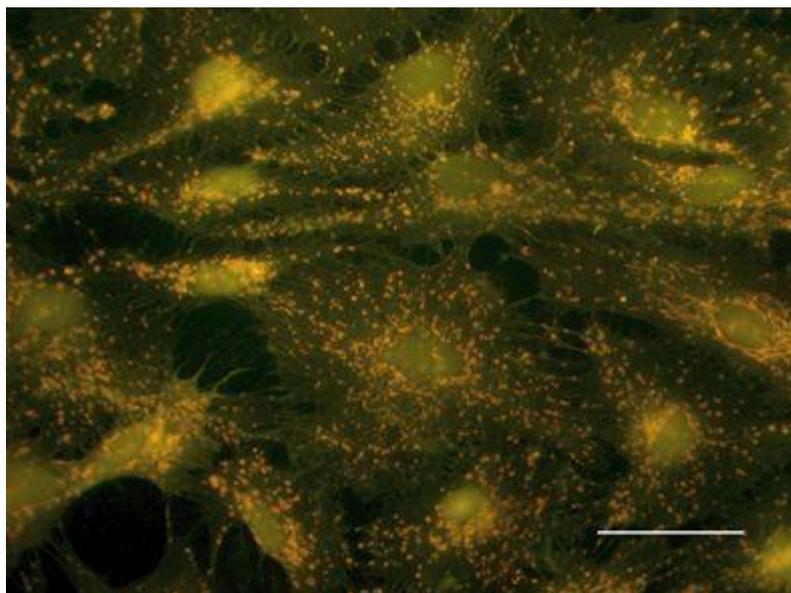


Figure 3: Fluorescence image of C6 glioblastoma cells labeled with QDs (incubation time = 3 minutes), obtained by single photon laser scanning confocal microscopy. Scale bar: 25 micrometers.

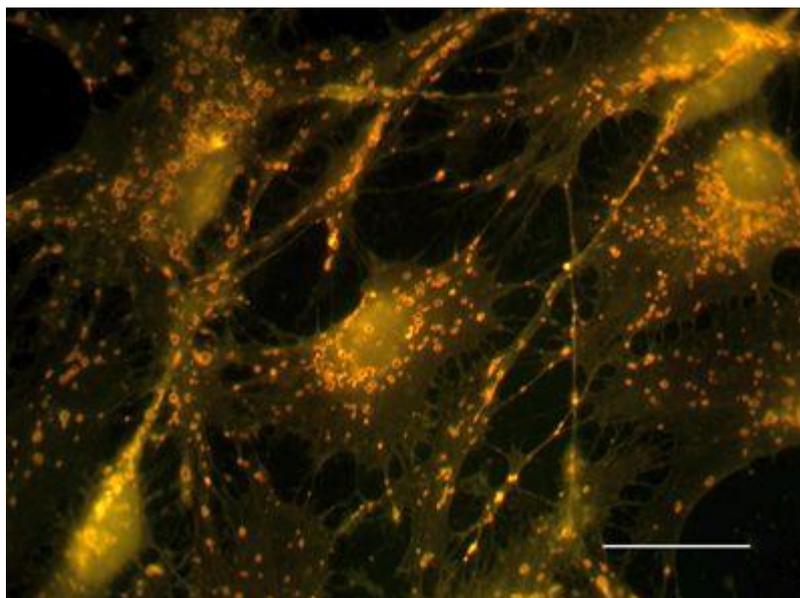


Figure 4: Fluorescence image of C6 glioblastoma cells labeled with QDs (incubation time = 7 days), obtained by single photon laser scanning confocal microscopy. Scale bar: 25 micrometers.

Contrast phase image obtained for the same times, 3 minutes and 7 days, are presented in Figure 5 left and right, respectively.

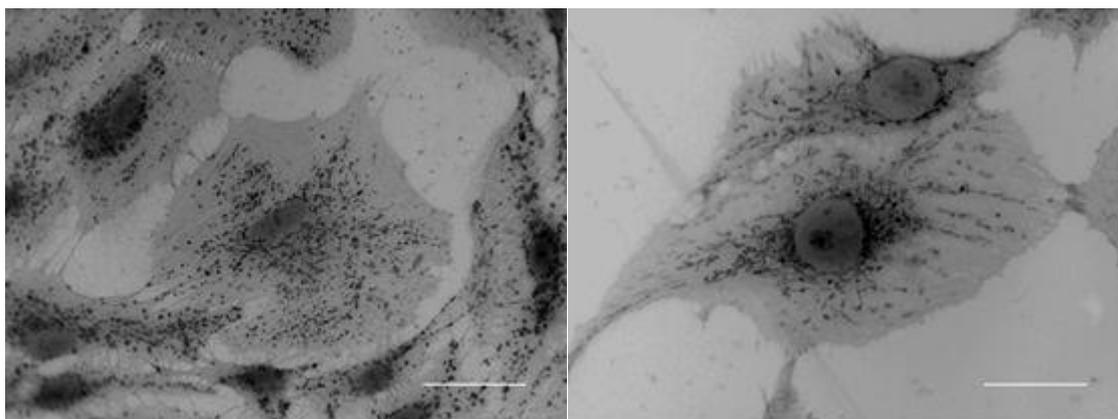


Figure 5: Phase contrast images of control (non-labelled) C6 glioblastoma cells (left: 3 minutes; right: 7 days). Scale bar: 25 micrometers.

In vivo toxicological with male Wistar rats studies via inhalation exposure have shown that the technically maximal possible concentration of free Cd ions corresponds to 0.52 mg Cd/m³. The results of the study indicate that there was no evidence that cadmium was translocated to the blood stream or to the central nervous system [20].

4. CONCLUSIONS

Biocompatible QDs were employed as fluorescent probes for long term *in vitro* labelling of living C6 glioblastoma cells. The endocytosis of the QDs and their apparent absence of deleterious effects to the cells may be associated with the efficiency of polyethylene-glycol (PEG) as functionalizing agent which seems to show an intense bond to the core/shell QDs' surface. PEGylated QDs, *a priori*, do not have the ability to target some specific site/organelle inside the cells. In C6 glioblastoma cells, QDs were quickly internalized, while in the case of healthy glial cells, they remained bound to cells' surface. In both cases,

presence of QDs did not negatively impact the *cell viability* and *normal cell dynamics*, which are central issues in the development of molecular imaging and drug/gene delivery systems. This study contributes as a prove of principle for a fluorescent nanoprobe-based assay, as the nanoprobe (QDs) employed have proved to be reliable for long term *in vitro* analysis and monitoring of cell mechanisms, such as migration of glioblastomas. This potentially paves the way for a better understanding of the gliomas, the most common kind of brain cancer.

DECLARATIONS

The experiments were performed with line cells commercially available. All the authors consent the publication and contributed equally to this work. There are no competing interests. CNPq (Grant 405030/2015-0) and FFG (Grant 861023) partially supported this work.

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